

Horticulture Innovation Australia

Final Report

ASDS8 – conference support for vegetable specialists

Dr Calum Wilson
Tasmanian Institute of Agriculture (TIA) - University of
Tas

Project Number: VG14700

VG14700

This project has been funded by Horticulture Innovation Australia Limited with co-investment from Australasian Soilborne Diseases Symposium and funds from the Australian Government.

Horticulture Innovation Australia Limited (HIA Ltd) makes no representations and expressly disclaims all warranties (to the extent permitted by law) about the accuracy, completeness, or currency of information in *ASDS8 – conference support for vegetable specialists*.

Reliance on any information provided by HIA Ltd is entirely at your own risk. HIA Ltd is not responsible for, and will not be liable for, any loss, damage, claim, expense, cost (including legal costs) or other liability arising in any way (including from HIA Ltd or any other person's negligence or otherwise) from your use or non-use of *ASDS8 – conference support for vegetable specialists*, or from reliance on information contained in the material or that HIA Ltd provides to you by any other means.

ISBN 0 7341 3563 7

Published and distributed by:
Horticulture Innovation Australia Limited
Level 8, 1 Chifley Square
Sydney NSW 2000
Tel: (02) 8295 2300
Fax: (02) 8295 2399

© Copyright 2015

CONTENTS

Media Summary	5
Technical Summary	6
ASDS8 conference and associated industry workshop	7
Introduction	7
ASDS8 conference	8
industry workshop.....	15
Conclusion.....	41
Technology Transfer	42
Scientific Conference papers.....	42
Industry & media publications	42
Industry & Peer presentations	42
Project Recommendations	43

TABLE OF FIGURES AND TABLES

Table 1	The ASDS8 conference program with presentation titles and authors	8
Fig 1	Group photo from the 8th Australasian Soilborne Diseases Symposium (8ASDS), November 2014 at The Hobart Function and Conference Centre Tasmania, Australia	14
Fig 2	Invitation letter to Industry Workshop	15

MEDIA SUMMARY

The 8th Australasian Soil-borne Diseases Symposium (8ASDS) was held in Hobart 10-13th November 2014. This is one of the premier meetings for scientists working with soil-borne disease, soil microbial ecology and plant-microbe interactions in Australasia. The conference attracted 85 scientists and other attendees from seven countries with research students networking strongly encouraged. The scientific programme encompassed topics including pesticides and soil amendments, integrated pest management, plant-pathogen interactions, biological control and disease suppression, resistance, pathogen ecology and epidemiology, and diagnostics and risk management.

Two international specialists in vegetable soil-borne disease R,D&E (Dr Alison Lees, The James Hutton Institute, UK; and Dr Krishna Subbarao, University of California, Davis, USA) were supported to attend the conference. They presented keynote addresses and participated in an industry workshop in NW Tasmania following the conference where they presented to an industry audience.

The ASDS8 meeting provided an important opportunity for scientists to share their latest results and thoughts, network and develop collaborations that will aid in the development of more efficient and effective R,D&E in the future. The industry workshop enabled industry to learn from the experiences of our international specialists in a non-academic setting providing time for questions and informal discussions.

TECHNICAL SUMMARY

The 8th Australasian Soil-borne Diseases Symposium (8ASDS) was held in Hobart 10-13th November 2014. This is one of the premier meetings for scientists working with soil-borne disease, soil microbial ecology and plant-microbe interactions in Australasia. The conference was opened by Tasmanian Liberal MP for Lyons, Mark Shelton and attracted 85 scientists and other attendees. This included participants from Australia, New Zealand, USA, UK, the Netherlands, Brazil, Tunisia, and Malaysia. A good contingent of research students were in attendance with opportunities for networking encouraged. The scientific programme encompassed topics including pesticides and soil amendments, integrated pest management, plant-pathogen interactions, biological control and disease suppression, resistance, pathogen ecology and epidemiology, and diagnostics and risk management.

The HIA grant was primarily used to bring two international specialists in vegetable soil-borne disease R,D&E to the conference. They presented keynote addresses at the conference and participated in an industry workshop in NW Tasmania (at the Forthside Vegetable Research Farm) following the conference where they presented to an industry audience.

The invited speakers were:

Dr Alison Lees (The James Hutton Institute, UK) - a specialist in molecular detection of soil-borne pathogens and development of markers for host resistance selection, who discussed the successes and failures of real-time PCR diagnostic assays as both a research tool and as a predictive tool for soil-borne diseases of potato

Dr Krishna Subbarao (University of California, Davis, USA) - specialist in ecology, epidemiology and integrated control of fungal diseases of vegetables, who discussed how consumer demand (human activities) can alter soil borne diseases, as illustrated with ongoing studies on *Sclerotinia sclerotiorum* and *Verticillium dahliae*.

The ASDS8 meeting provided an important opportunity for scientists to share their latest results and thoughts, network and develop collaborations that will aid in the development of more efficient and effective R,D&E in the future.

The industry workshop enabled industry to learn from the experiences of our international specialists in a non-academic setting providing time for questions and informal discussions.

ASDS8 CONFERENCE AND ASSOCIATED INDUSTRY WORKSHOP

INTRODUCTION

The 8th Australasian Soil-borne Diseases Symposium (8ASDS) was held in Hobart 10-13th November 2014. This is one of the premier meetings for scientists working with soil-borne disease, soil microbial ecology and plant-microbe interactions in Australasia. The conference attracted 85 scientists and other attendees. This included participants from Australia, New Zealand, USA, UK, the Netherlands, Brazil, Tunisia, and Malaysia. A good contingent of research students were in attendance with opportunities for networking encouraged.

The HIA project had the following aims:

1. Assist in facilitating running of the ASDS8 conference.
2. Provide resources to invite two keynote speakers (experts in vegetable pathology) from the UK and the USA to the meetings.
3. Support an industry session where keynote speakers will present to an industry audience.
4. Support a prize for the best student presentation.

The HIA grant was used to bring two international specialists in vegetable soil-borne disease R, D & E to the conference. They presented keynote addresses at the conference and participated in an industry workshop in NW Tasmania (at the Forthside Vegetable Research Farm) following the conference where they presented to an industry audience.

The invited speakers were:

Dr Alison Lees (The James Hutton Institute, UK) - a specialist in molecular detection of soil-borne pathogens and development of markers for host resistance selection.

Dr Krishna Subbarao (University of California, Davis, USA) - specialist in ecology, epidemiology and integrated control of fungal diseases of vegetables.

The ASDS8 meeting provided an important opportunity for scientists to share their latest results and thoughts, network and develop collaborations that will aid in the development of more efficient and effective R, D&E in the future.

Quality student presentations were delivered throughout the conference. Natalia Cripps-Guazzone (Lincoln, University, New Zealand) was awarded a prize of \$200 for best student presentation.

Immediately following the conference an industry workshop was held which enabled vegetable industry members to learn from the experiences of our international specialists in a non-academic setting providing time for questions and informal discussions.

ASDS8 CONFERENCE

The ASDS8 meeting provided an important opportunity for scientists to share their latest results and thoughts, network and develop collaborations that will aid in the development of more efficient and effective R, D&E in the future (Fig 1.).

The HIA grant was used to bring two international specialists in vegetable soil-borne disease R, D & E to the conference. They presented keynote addresses at the conference and participated in an industry workshop in NW Tasmania (at the Forthside Vegetable Research Farm) following the conference where they presented to an industry audience.

The invited speakers were:

Dr Alison Lees (The James Hutton Institute, UK) - a specialists in molecular detection of soil-borne pathogens and development of markers for host resistance selection.

Dr Krishna Subbarao (University of California, Davis, USA) - specialist in ecology, epidemiology and integrated control of fungal diseases of vegetables.

Student prize:

Quality student presentations were delivered throughout the conference. Natalia Cripps-Guazzone (Lincoln, University, New Zealand) was awarded a prize of \$200 for best student presentation.

Table 1: the ASDS8 conference program with presentation titles and authors

SESSION 1 - PESTICIDES AND SOIL AMENDMENTS

EVALUATION OF COMPOST TEAS FROM MATURED AGRICULTURAL WASTE, VERMICOMPOST AND SOLID MUNICIPAL WASTE COMPOSTS TO SUPPRESS BACTERIAL WILT OF POTATO

W.K. Mengesha, S. Powell, K. Evans and K.M. Barry

TARGETED APPLICATION IMPROVES EFFICACY OF IN-CROP FUNGICIDES AGAINST CROWN ROT IN WHEAT

S. Simpfendorfer, F. Fensbo and R. Shapland

CRITICAL PHOSPHOROUS ACID LEVELS TO MANAGE PHYTOPHTHORA ROOT ROT OF AVOCADO

D.J. Armour and E.K. Dann

RHIZOCTONIA CONTROL IMPROVED BY LIQUID BANDING OF FUNGICIDES

P. Bogacki, J. Desbiolles, R. Correll, D. Hüberli, W. MacLeod and A. McKay

NEMATOSTATIC NATURAL PRODUCTS FROM UNNATURAL SOURCES

A.G.W. Murray, M. R. Rocha, C.B. Soll, T.L. Rocha and M.F. Grossi de Sa

SESSION 2 - INTEGRATED PEST MANAGEMENT

KEYNOTE: HUMAN ACTIVITIES AND SOILBORNE DISEASES: CONTEMPORARY EXPLORATIONS

K.V. Subbarao

THE CUP IS HALF FULL: SUCCESSFUL MANAGEMENT OF ROOT-LESION NEMATODES IN THE RAINFED SUBTROPICAL WHEAT REGION OF AUSTRALIA

K.J. Owen, T.G. Clewett and J.P. Thompson

COMPARISON OF CHEMICAL, BIOLOGICAL, AND CULTURAL TOOLS TO MANAGE SCLEROTINIA DROP OF LETTUCE

M.E. Matheron and M. Porchas

PHYTOPHTHORA CINNAMOMI IN MACADAMIA: ITS IMPACT AND STRATEGIES FOR SUSTAINABLE MANAGEMENT

O.A. Akinsanmi

SESSION 3 - PLANT/PATHOGEN INTERACTIONS

INVESTIGATING SOILBORNE NECTRIACEOUS FUNGI ASSOCIATED WITH BLACK ROOT ROT IN AVOCADO

L.E. Parkinson, A.R. McTaggart, R.G. Shivas, and E. K. Dann

INFECTION AND COLONISATION OF POTATO PLANTS BY COLLETOTRICHUM COCCODES IN AUSTRALIA

J. Chang, R.F. de Boer, P.W. Crous and P.W.J. Taylor

POTATO ROOT EXUDATES STIMULATE ZOOSPORE RELEASE OF SPONGOSPORA SUBTERRANEA

M. Balendres, D. Nichols, R. Tegg and C. Wilson

STRIGOLACTONES, A NEW HORMONE GROUP WITH ROLES IN PLANT-MICROBE INTERACTIONS

S.N. Blake, K.M. Barry, B.Reid and E. Foo

FUSARIUM PSEUDOGRAMINEARUM CROWN ROT: GROWTH PATTERNS IN PLANTA

N.L. Knight and M.W. Sutherland

POSTER SESSION 1

SOMATIC CELL SELECTION AS A TOOL FOR DEVELOPING RESISTANCE AGAINST SOIL-BORNE POTATO PATHOGEN, SPONGOSPORA SUBTERRANEA

M. Balendres, R. Tegg and C. Wilson

EFFECT OF CROWN INFECTION BY STAGONOSPOROPSIS TANACETI ON GROWTH AND DEVELOPMENT OF PYRETHRUM

M.A.H.B. Bhuiyan and P.W.J. Taylor

CALCULATING HEAT HOURS FOR MAXIMUM PRODUCTION OF EGGS FOR 3 SPECIES OF MELOIDOGYNE

J.A. Cobon, W. O'Neill and T. Shuey

PRATYLENCHUS PENETRANS, A DEVASTATING PEST IN WESTERN AUSTRALIAN BROADACRE CROPS

S.J. Collins, C.J. Wilkinson, H.F. Hunter, L. DeBrincat and S.J. Kelly

RESISTANCE OF FIELD PEAS TO *PRATYLENCHUS THORNEI*

J.P. Fanning, P. R. Kennedy, B.J. Gogel and G.J. Hollaway

A NATIONAL APPROACH TO THE MANAGEMENT OF ROOT LESION NEMATODES IN AUSTRALIAN FIELD CROPS

G. Hollaway, S. Collins, A. McKay, K. Owen, S. Simpfendorfer and S. Taylor

IT'S A MYSTERY: WHY IS THERE DISEASE PRESENT IN CEREAL ROOTS IN THE ABSENCE OF PATHOGEN DNA IN THE SOIL?

D. Hüberli, S. Collins, M. Connor, W. MacLeod, R. Correll, and A. McKay

NEW ROLES FOR THE PLANT HORMONES STRIGOLACTONES, GIBBERELLINS AND AUXIN INFORMATION OF BENEFICIAL SYMBIOSIS WITH ARBUSCULAR MYCORRHIZAL FUNGI

C.J. Hugill, L.J. Quittenden, J.J. Ross, J.B. Reid and E. Foo

A QUALITATIVE PCR FOR DETECTION OF ASCOSPORIC INOCULUM OF *SCLEROTINIA SCLEROTIUM*

S.J. Jones, S. Pilkington, D.H. Gent, F.S. Hay and S.J. Pethybridge

OCCURRENCE OF ROOT-LESION NEMATODES IN CENTRAL QUEENSLAND, AUSTRALIA

N.A. Robinson, J.G. Sheedy, T.G. Clewett, M. Conway and J.P. Thompson

SPONGOSPORA SUBTERRANEA ROOT INFECTION ASSESSED IN SIX POTATO CULTIVARS FOR RELATIVE SUSCEPTIBILITY TO POWDERY SCAB

A. Agarwal, T.J. Wiechel, F. Richardson, D. Auer and R.F. de Boer

EVALUATING SALICYLIC ACID TO INDUCE PLANT INNATE IMMUNITY AGAINST *SPONGOSPORA SUBTERRANEA* ROOT INFECTION OF POTATO

J.T. Lamattina, A. Agarwal, J. Rookes, R. F. de Boer, D. Auer and T.J. Wiechel

SESSION 4 - BIO-CONTROL

A SIMULATORY STUDY LINKING CELLULASE ACTIVITIES BY ENDOPHYTIC BIOCONTROL AGENTS TO THEIR COLONIZATION EXTENT 20

H.S. Yoo and A.S.Y. Ting

ENDOPHYTIC ACTINOBACTERIA THAT ENHANCE RHIZOBIAL FUNCTION AND CONTROL ROOT DISEASES IN LUCERNE

X.H. Le, C.M.M. Franco and R. Ballard

BIOCONTROL OF WHEAT TAKE-ALL: SEED COATING WITH A FOCUS ON *TRICHODERMA*

D. Bienkowski, E. Hicks, M. Braithwaite and R.E. Falloon

BIOLOGICAL CONTROL OF PASTURE BARE-PATCH DISEASE WITH *TRICHODERMA* BIO-INOCULANTS

D.R.W. Kandula, A. Stewart, E. Duerr, J.G. Hampton and D. Gale

BIOLOGICAL CONTROL OF ONION WHITE ROT USING *TRICHODERMA* ENRICHED COMPOSTS

D.A. Metcalf, C.R. Wilson, J.C. Dennis and T. Groom

RHIZOSPHERE COMPETENCE IN *TRICHODERMA* SPECIES

N. Cripps-Guazzone, E.E. Jones, L.M. Condrón, R. Hill, A. Stewart and H.J. Ridgway

SESSION 5 - MANAGEMENT OF SOILBORNE DISEASES OF CEREALS - DAVID ROGET REMEMBERED

MANAGEMENT OF TAKE-ALL IN WHEAT

Matthew G. Cromey, S.L. Bithell, S.F. Chng, R.F. van Toor and A.C. McKay

AGRONOMIC MANAGEMENT OF SOIL-BORNE DISEASES

John Kirkegaard

PLANT RESISTANCE TO SOILBORNE DISEASES

Karam Singh

MOLECULAR APPROACH TO MICROBIAL ECOLOGY UNRAVELS SOILBORNE DISEASE SUPPRESSIVE COMMUNITIES

Vadakattu V.S.R. Gupta, C.R. Penton, J.M. Tiedje, P. Greenfield, S.N. Neate, M. Gillings, K. Ophel-Keller, P. Harvey and D.K. Roget

SESSION 6 - IPM/CULTURAL CONTROL

KEYNOTE: SOIL-BORNE DISEASES OF POTATO: EPIDEMIOLOGY, PREDICTION AND MANAGEMENT

Alison K. Lees, J.L. Brierley, V.C. Blok and R. Neilson

SOILBORNE DISEASES IN SEED POTATO CERTIFICATION

N.S. Crump

GENETIC, GENOMIC AND BIOLOGICAL CONTROL STRATEGIES TO MANAGE *FUSARIUM* CROWN ROT OF WHEAT

K. Kazan, D.M. Gardiner, F. Obanor, M.M. Roper, P. Harvey

USE OF VEGETATED GROUNDCOVERS CAN SUPPRESS PANAMA DISEASE IN BANANAS

A.B. Pattison, T.L. Kukulies, E. Rames, W. O'Neill, J.A. Cobon, C. Wright and A. Molina

SESSION 7 - RESISTANCE AND CHEMICAL CONTROL

ENHANCING RESISTANCE TO SOILBORNE PATHOGENS OF SUGARCANE THROUGH INTROGRESSION BREEDING

S.A. Bhuiyan, B.J. Croft, P. Jackson, G.R. Stirling, R. Magarey, E. Wong and J. Bull

RESISTANCE AND TOLERANCE OF CEREAL CULTIVARS TO ROOT LESION NEMATODE *PRATYLENCHUS NEGLECTUS* AND *P. THORNEI* IN SOUTH-EASTERN AUSTRALIA

A. McKay, K. Linsell, R. Davey, J. Fanning, P. Bogacki, B. Gogel and G. Hollaway

CRITICAL CHALLENGES FOR THE PHASE-OUT OF METHYL BROMIDE IN THE AUSTRALIAN STRAWBERRY INDUSTRY

S. Mattner, I. Porter, P. Merriman and M. Milinkovic

NATIONAL STRAWBERRY INDUSTRY THREATENED

P. Merriman, S. Mattner G. Weda and M. Milinkovic

THE IMPACT OF METHYL BROMIDE PHASE OUT ON NEW TECHNOLOGIES FOR CONTROL OF SOILBORNE DISEASES, BIOSECURITY AND THE OZONE LAYER

I.J. Porter, S.W. Mattner and D.A.Riches

POSTER SESSION 2

COMBINING MICROBIAL ENEMIES FOR ENHANCED CONTROL OF ROOT LESION NEMATODES

K. Crampton, E. Adorada, C. Lisle, M. Hodda and G. Ash

INFLUENCE OF ELEVATED SOIL SULPHUR, ZINC AND IRON AND A FUNGICIDE ON POWDERY SCAB DISEASE AND YIELD OF TWO POTATO CULTIVARS: A FIVE YEAR STUDY

T.J. Wiechel, A. Agarwal, D.P.F. Auer, F. Richardson, M. Wardzynski, J. Edwards and R.F. de Boer

EFFICACY OF SOIL AMENDMENT TO CONTROL VERTICILLIUM WILT DISEASE

V.N. Dharjono, N.S. Crump, T. Wiechel and P.W.J. Taylor

UREA IN COMBINATION WITH BURIAL REDUCES THE PRODUCTION OF PSEUDOTHECIA ON *LEPTOSPHAERIA MACULANS* INFECTED STUBBLE

S. Lob, H.J. Ridgway, S.A. Wakelin, M.V. Jaspers and E.E. Jones

MARKER-ASSISTED SELECTION OF BACTERIAL BIO-CONTROL AGENTS FOR THE CONTROL OF SCLEROTINIA DISEASES

M.M. Kamal, K.D. Lindbeck, S. Savocchia, G.J. Ash and B.B. McSpadden Gardener

RECAPTURED QUARANTINE METHYL BROMIDE: AN ALTERNATIVE TO METHYL BROMIDE FOR SOIL DISINFESTATION?

S. Mattner, I. Porter, J. Falco and W. Grullemans

YIELD AND QUALITY EFFECTS OF POTATO FROM *MELOIDOGYNE FALLAX*

M. Rettke, B.H. Hall and G. Walker

DEVELOPING BIOCONTROL AGENTS AGAINST *FUSARIUM* CROWN ROT IN WHEAT

M.M. Roper, K. Kazan, P.R. Harvey, F. Obanor, R. Poels, C.A. Myers, R. Warren, B. Stummer and R. Sabburg

BIODISCOVERY OF ANTIMICROBIAL COMPOUNDS FROM PLANT GROWTH-PROMOTING RHIZOBACTERIA (PGPR) AND THEIR ROLE AGAINST *PHYTOPHTHORA CINNAMOMI*

A.R. Rosli, L. Carvalhais, Z. Khalil, R. Capon and P.M. Schenk

EVIDENCE OF BIOLOGICAL SUPPRESSION OF *PRATYLENCHUS THORNEI* IN THE NORTHERN GRAIN REGION OF AUSTRALIA

N.P. Seymour, G.R. Stirling and Y. Li

TIME OF PLANTING WHEAT CAN MODIFY YIELD LOSS FROM ROOT-LESION NEMATODE (*PRATYLENCHUS THORNEI*) IN SUB-TROPICAL AUSTRALIA

J.P. Thompson, T.G. Clewett and M.M. O'Reilly

YIELD LOSSES IN WESTERN AUSTRALIAN CEREAL CROPS CAUSED BY ROOT LESION NEMATODES

C.J. Wilkinson and S.J. Collins

SESSION 8 - DISEASE SUPPRESSION

METABOLOMIC APPROACHES FOR THE DISCRIMINATION OF DISEASE SUPPRESSIVE SOILS FOR
RHIZOCTONIA SOLANI AG8

H.L. Hayden, S. Rochfort, V. Ezernieks and P.M. Mele

INTERACTION BETWEEN ROOTSTOCK CULTIVAR AND AMF SPECIES INFLUENCES SUSCEPTIBILITY TO
ILYONECTRIA SPP. INFECTION

E.E. Jones, S.E.H. Hammond, D.S. Brown, C. Blond and H.J. Ridgway

FREE LIVING NEMATODES AS INDICATORS OF THE BIOLOGICAL STATUS OF AUSTRALIAN CEREAL SOILS

K. Linsell, A. Stirling, D. Hartley, Herdina, A. Cheshire, J. Nobbs, A. McKay, G. Stirling and K. OphelKeller

BIOLOGICAL SUPPRESSION OF *PRATYLENCHUS NEGLECTUS* AND *P. THORNEI* IN WESTERN AUSTRALIAN AND
SOUTH AUSTRALIAN GRAIN-GROWING SOILS

G.R. Stirling, K. Linsell and L. Martin

MICROBIAL SUPPRESSION OF *RHIZOCTONIA* ROOT ROT OF WHEAT

S.J. Barnett, S. Zhao, R. Ballard and C. Franco

PUTATIVE SUPPRESSION OF COMMON SCAB OF POTATO IN A NATURAL FIELD SOIL

D.P.F. Auer, T.J. Wiechel, D. Johnston-Monje, J.M. Park, G. Lazarovits and R. de Boer

SESSION 9 - PATHOGEN ECOLOGY AND EPIDEMIOLOGY

PLANT-SOIL INTERACTIONS COULD CONTRIBUTE TO DIEBACK IN A WIDESPREAD INVASIVE EXOTIC PLANT,
PARKINSONIA ACULEATA

A.K.H. Raghavendra, P.H. Thrall, A. Bissett, T. Steinrucken and R.D. van Klinken

DELAYING *SPONGOSPORA* INDUCED ROOT INFECTION IN POTATO ROOTS BY ALTERING INOCULATION
DATES AND CHEMICAL TREATMENT

T. Thangavel, R.S. Tegg and C.R. Wilson

SOILBORNE DISEASE POTENTIAL IN VICTORIAN POTATO FIELDS

T.J. Wiechel, N.S. Crump, A. McKay and K. OphelKeller

IDENTIFICATION AND CHARACTERISATION OF GENETIC MARKERS IN PULSE CROPS THAT PROMOTE DISEASE
RESISTANCE WITHOUT COMPROMISING SYMBIOTIC INTERACTIONS

J.M. Plett, K.L. Plett, J.R. Powell and I.C. Anderson

INVESTIGATING THE BIOLOGY, EPIDEMIOLOGY AND MANAGEMENT OF *NEOCOSMOSPORA* ROOT ROT OF
PEANUT IN AUSTRALIA

K.M. Wenham, V.J. Galea, M.J. Ryley, G. Wright and W. Bryden.

MANAGEMENT PRACTICES INFLUENCE *RHIZOCTONIA SOLANI* AG8 INOCULUM AND DISEASE IMPACTS IN
CEREAL CROPS

V.V.S.R. Gupta, A. McKay, K. Ophel-Keller, J. Kirkegaard, N. Wilhelm and D.K. Roget

SESSION 10 - DIAGNOSTICS AND RISK MANAGEMENT

INTRASPECIFIC VARIATION AMONG ISOLATES OF *PYTHIOGETON RAMOSUM* ISOLATED FROM SOFT ROT
DISEASE GINGER IN QUEENSLAND, AUSTRALIA

D.P. Le, M.K. Smith and E.A.B. Aitken

ADDITION OF STUBBLE TO PREDICTA B SOIL SAMPLES ENHANCES THE DETECTION OF CROWN ROT
S.A. Rowe and A. McKay

CONTRIBUTION OF SPATIAL ANALYSIS TO UNDERSTANDING RISK AND MORTALITY IN *ACACIA MANGIUM* IN
CENTRAL SUMATRA
A. Nawari, M.A. Hardie, C. Mohammed, D. Mendham, M. Glen, M.P. Periasamy and A.Gafur

IDENTIFICATION OF FUNGAL PATHOGENS ASSOCIATED WITH SLOW PLANT LOSS AFTER SINGLE HARVEST
(SPLASH) SYNDROME IN PYRETHRUM FIELDS OF TASMANIA
A. Moslemj, T. Groom and P.W.J. Taylor

PRE-SOWING *PRATYLENCHUS THORNEI* DENSITY CAN BE USED TO DETERMINE YIELD LOSS RISK
IN CEREAL CROPS GROWN IN VICTORIA
J. Fanning, A. McKay, C. Forknall, and G. Hollaway



Fig 1: Group photo from the 8th Australasian Soilborne Diseases Symposium (8ASDS), November 2014 at The Hobart Function and Conference Centre Tasmania, Australia

INDUSTRY WORKSHOP

The industry workshop was held on the morning of Friday 14th November at the Forthside Vegetable research Farm (Fig 2.).

Presentations were given by the two international speakers to an audience of around 30-40 industry representatives.

The presentations were followed with a question session, and then lunch provided during which further interaction with the international guests was encouraged. Prof Subbarao was interviewed by AUSVEG representatives and featured within the following Vegetable Australia magazine highlighting his work managing *Verticillium dahliae* and *Sclerotinia* in brassica and lettuce crops, in particular about how changing farm management systems has altered disease incidence.

Vegetable growers' workshop

14th November 10.00 AM

A workshop for vegetable growers and industry will be hosted by the Tasmanian Institute of Agriculture (TIA) at Forthside Vegetable Research Facility, 125 Forthside Rd, Forth, TAS.

The workshop will feature discussions about diseases relevant to local growers and will include presentations from Alison Lees from the Hutton Institute in the UK and Krishna Subbarao from University of California – Davis. The workshop will include formal presentations and open discussions with the international speakers and other TIA plant pathologists (Dr Calum Wilson).



Dr Alison Lees: a plant pathologist will discuss diagnosis and management of soil-borne potato and vegetable diseases, based on her experience in the UK.



Dr Krishna Subbarao: an extension plant pathologist will talk briefly about vegetables in coastal California and then on 'Mechanisms of Broccoli-mediated soilborne pathogen and disease suppression in vegetable cropping systems'.

A short farm walk looking at some of TIA's latest research projects is also included.

The workshop runs from 10:00am and finishes with a free lunch at 12:30pm (no need to RSVP).

For more information please contact Robert Tegg via email on Robert.Tegg@utas.edu.au.

The workshop is part of a HAL supported project funded through the National Vegetable Levy.



Fig 2: Invitation letter to Industry Workshop

A copy of slides of these two presentations are provided here.

Dr Alison Lee's Presentation at Industry Workshop

Potato (& veg) diseases: UK research

Alison Lees & Jennie Brierley, JHI.



Our sites

Craigiebuckler, Aberdeen
Laboratories



Invergowrie, Dundee
Laboratories, glasshouses and arable land



Hartwood Research Station, Lanarkshire
350ha rotational and permanent
grassland, moor and woodland



Glensaugh, Kincardineshire
865ha rotational grassland,
permanent pasture, heather moor
and peat



Balruddery Farm, Angus
Arable farm
Centre for Sustainable Cropping

UK vegetable sector

- UK Field Vegetable sector valued at ~£690 million. Covers > 130,000 ha, contributes > 30% of the total levy income.
 - Primary R&D focus has been crop protection (75% of the research budget).
1. Availability of adequate and sustainable **crop protection measures**.
 - EAMU (formerly SOLA) programme of paramount importance. Maintain anti-resistance strategies. New chemistry, non-chemical approaches, towards pesticide-free produce.
 2. Increasing returns on investment through the **efficient use of resources**
 - Improve farm efficiency, increased automation, precision agronomy, water, soil and waste management.
 3. Supply consistent **quality product and continuity of supply**.
 - Breeding for P&D resistance and quality attributes, improving storage regimes, optimising nutrition and harvest scheduling.
 4. **Co-ordinated approach to R&D and technology transfer to farm level**.
 - Ensure advances are taken up and used for the benefit of the UK industry as a whole. Improve links with allied levy bodies, DEFRA and consultants.
 5. Information to **promote consumption of home grown vegetables**.
 - Heightening the awareness of consumers to the health benefits of vegetables and encouraging consumption.



HDC project FV 417 Crops and pathogens
Nicola Holden



- Cabbage – blight / black rot (*Xanthomonas*)
- Broccoli – head rot (*Pseudomonas; Pectobacterium*)
- Radish – leaf blight (*Pseudomonas*)
- Brussels sprouts – light leaf spot (*Pyrenopeziza*)
- Red onion – soft rot (*Burkholderia*)

Aims



- No real treatment options for bacteria, except for copper oxychloride, which is toxic to the environment.
- Aim to promote plant defence response to help reduce bacterial load on plants.
- This project used treatments either already commercially available or with a good chance at being available soon (rather than 'raw' chemicals).

Are elicitors commercially available?



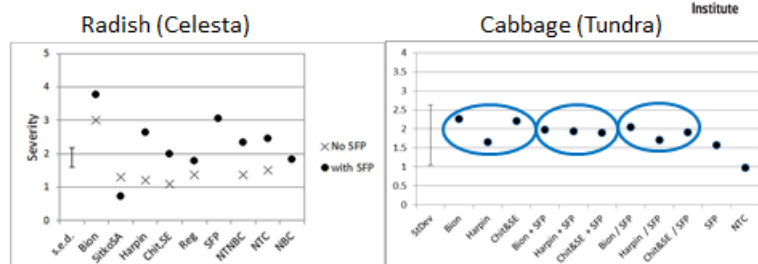
Product	Elicitor activity	Current use	Prospects
Amistar	Strobilurin	Brassicas: White Blister, Ring Spot, Alternaria Onion and Radish: Downy Mildew	Good
Signum	Strobilurin	Brussels sprouts, cabbage, broccoli and radish: Downy mildew	Good
Bion (Syngenta)	ASM: SA mimic	Actiguard (US) Label approved for various including Brassicas for Xanthomonas (black rot)	Good - Fair
Sitko-SA (Growth Products USA)	SA + phosphite	Sold as a fertilizer in the USA. Not currently sold in UK, but can be shipped.	Good - Fair
Softguard (Travena, UK)	Chitosan	Sold as a plant health-care or growth promoter product (fertiliser) in the UK	Good
Algal 600 products (Travena, UK)	Seaweed extracts, laminarin	Sold as a nutritional supplement in the UK, often combined with Softguard.	Good
Harpin (Plant Health Care, USA)	Secreted protein	Sold as a plant health promoter, available in the UK via Plant Health Care, UK office.	Good - Fair
Regalia / Reysa, Milsana (Syngenta)	Knotweed extract	To be marketed in Europe by Syngenta. Used on range of crops to control wide range of pathogens.	Good - Fair

Testing elicitor treatments for bacterial disease of Brassica



- Variety differences (same pathogen diff. variety)
- Species differences (same plant diff. pathogen)

Interactions with conventional fungicides

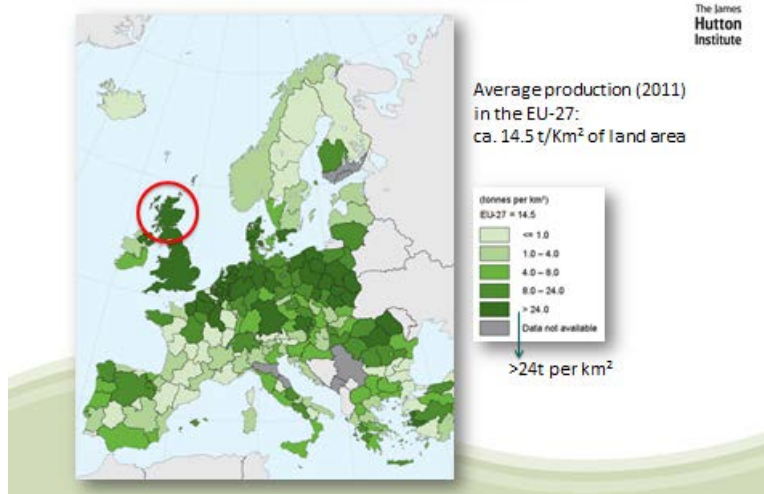


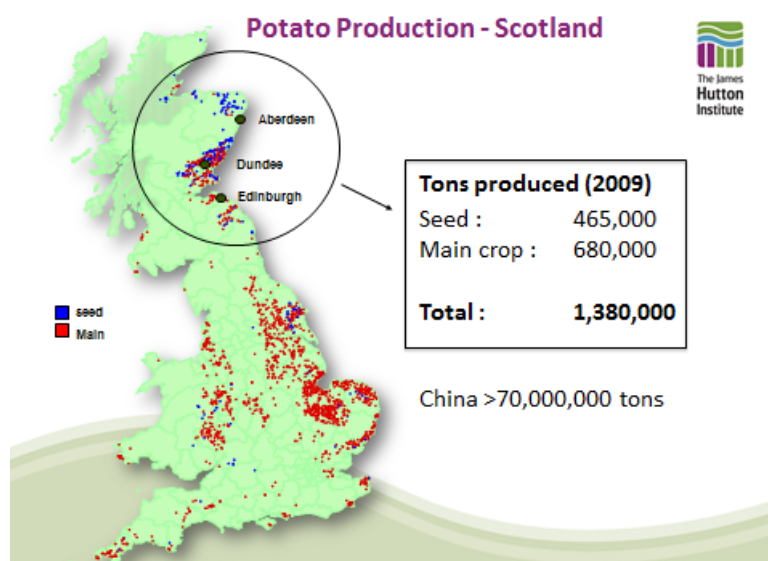
In some instances elicitors with SFP increased disease severity, e.g. Harpin on Radish.

Conclusion:

- Elicitors can work, but in a very 'system' dependent manner
- Can be applied, but in an informed and targeted manner

Potato production - Scotland



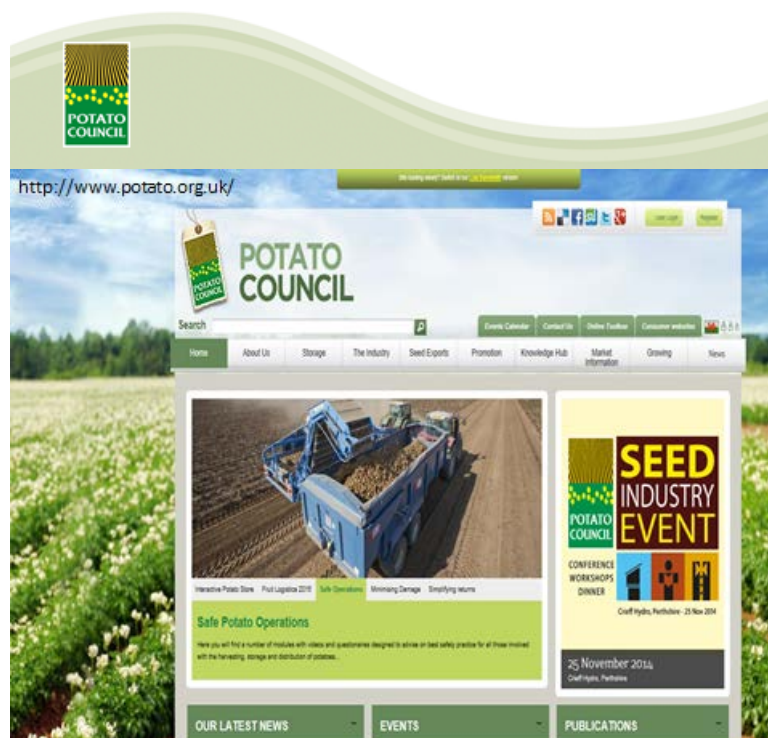


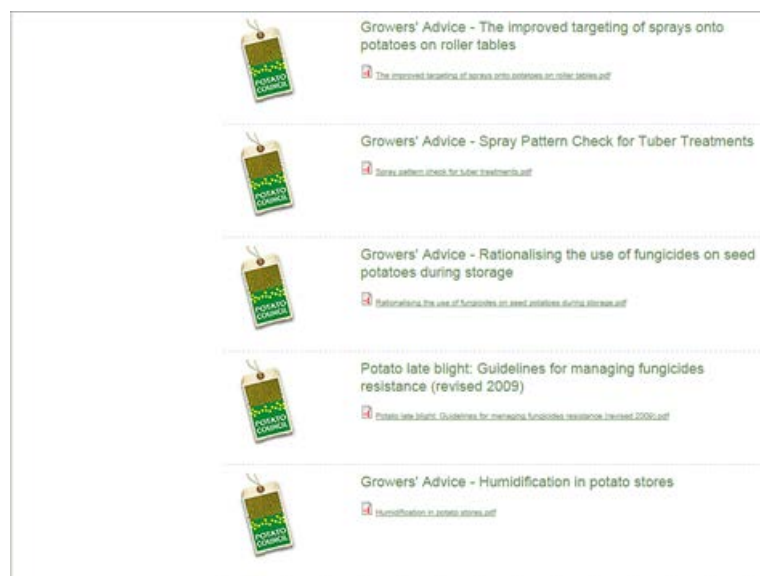
Potato



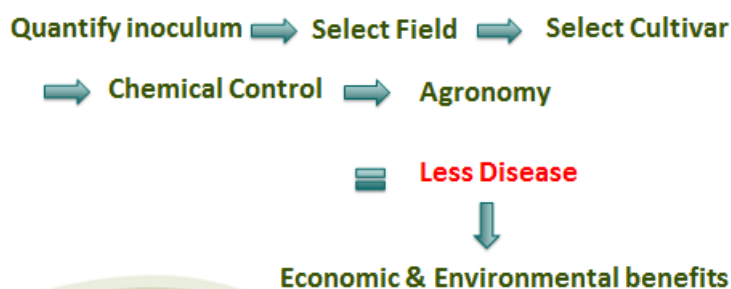
Potato Council

- Projects aim to improve marketable yield, reduce defects, reduce input costs.
- Currently around 30 projects
 - total value of ~£10m (£1m is levy payer funding, remainder coming from other sponsors such as Scottish Government, Defra, Technology Strategy Board, BBSRC and potato industry businesses).
- 5 key areas: pest; disease; agronomy; storage; and consumer-related research.
- Projects are carried out at research centres throughout the UK and they involve collaborators in Australia, New Zealand, Switzerland and South Africa.





Disease Management – soil-borne diseases



The role of diagnostics in the management of soil-borne potato diseases: considerations



- Quantitative assay – which pathogen and how much?
- Sampling strategy – can we find the pathogen in the field?
- What do the results mean?
 - Inoculum thresholds for risk
 - Spread of individual diseases
 - Effect of environment on disease risk
 - Available control measures



Quantitative assay

- Specific real-time PCR assays available for most potato pathogens
- Able to detect very low levels of pathogen DNA in plant, tuber, soil and water samples.
- DNA of a specific pathogen is quantifiable

Sampling strategy



Soil sampling strategy underpins the reliability of soil testing.

- must be representative of field scale
- must be practical (sampling and processing time and cost)
- based on “old” PCN sampling strategy.
- Sampling area: 4ha or less (divide larger fields)
- Sample size: 1Kg for standard testing
- Sampling points: 100 x 10g samples(0-15 cm depth)
- Sampling pattern: W pattern.

Brierley *et al.*, 2009. Quantifying potato pathogen DNA in soil.
Applied Soil Ecology 41, 234-8.

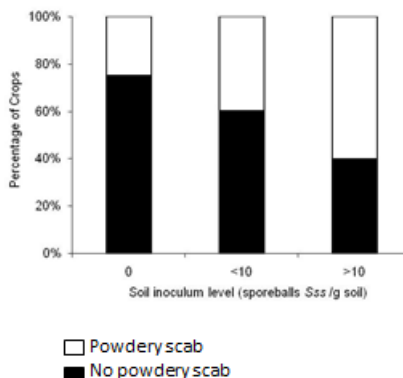
What do the results mean?

- **Inoculum Thresholds**



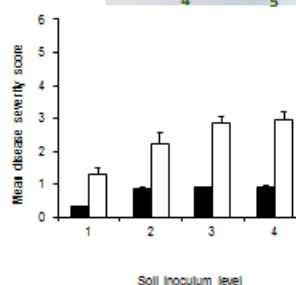
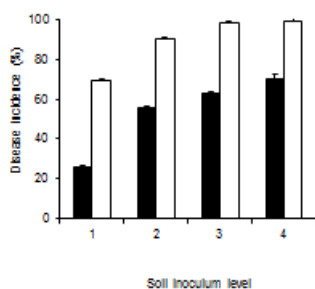
Monitoring of commercial potato fields

In commercial crops the risk of progeny tubers having powdery scab increased from 25% to 65% based on pre-plant levels of soil inoculum.
 Seed-borne inoculum responsible for disease where no soil-borne inoculum detected



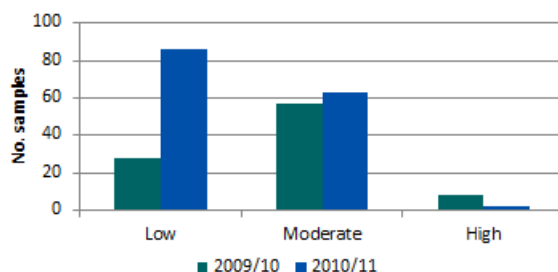
Inoculum levels: field trial results, powdery scab

- Level 4 = 40-50 sporeballs/g soil.
- □ = Agria ■ = Nicola



Brierley et al (2013 in press). Relationship between *Spongospora subterranea* f. sp. *subterranea* soil inoculum level, host resistance and powdery scab on potato tubers in the field. *Plant Pathology*.

Commercial soil diagnostic testing - powdery scab SRUC 2009/10 and 2010/11



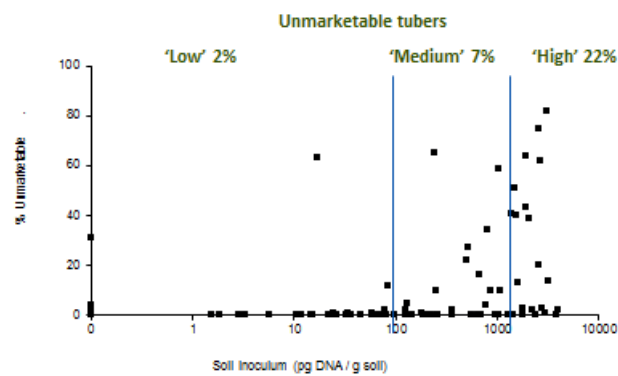
Conclusions

- Three years of field trial results show that the level of soil inoculum significantly affects amount of powdery scab on progeny tubers.
- Evidence towards use of diagnostic test for field selection



Colletotrichum coccodes - black dot soil inoculum

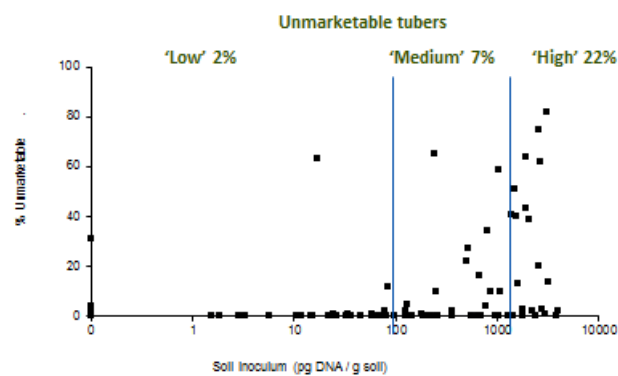
120 commercial fields x 3 years



Lees et al (2010). Relative importance of seed-tuber and soil-borne inoculum in causing black dot disease of potato. *Plant Pathology* 59, 693-702.

Colletotrichum coccodes - black dot soil inoculum

120 commercial fields x 3 years



Lees et al (2010). Relative importance of seed-tuber and soil-borne inoculum in causing black dot disease of potato. *Plant Pathology* 59, 693-702.

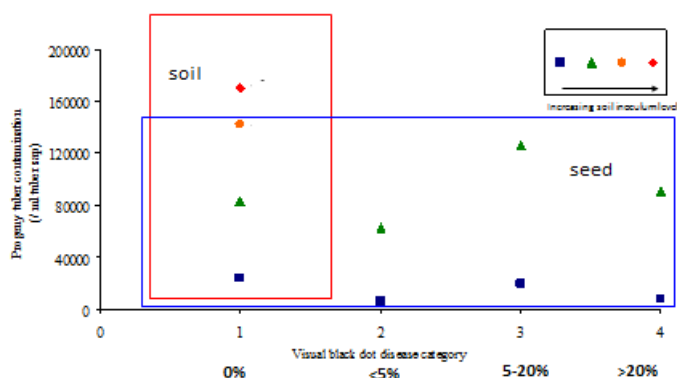
What do the results mean?

• Epidemiology of individual diseases

- Where does the inoculum come from ?
- When does infection takes place?
- What factors affect the development of symptoms?

Where does the inoculum come from?

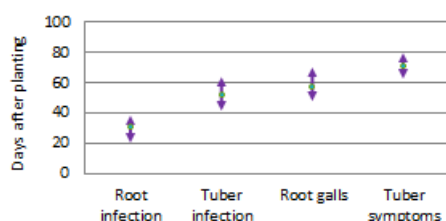
Effect of seed- and soil-borne inoculum on progeny tuber contamination by *C. coccodes*



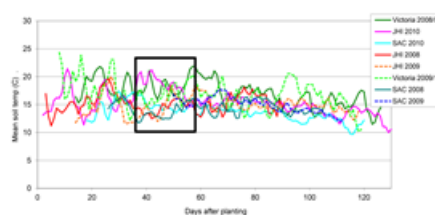
Loos et al., 2010. Relative importance of seed-tuber and soil-borne inoculum in causing black dot disease of potato. *Plant Pathology* 55, 695-702

Powdery scab infection and symptoms

New information on infection and disease development
Trials in Scotland, Victoria, Tasmania

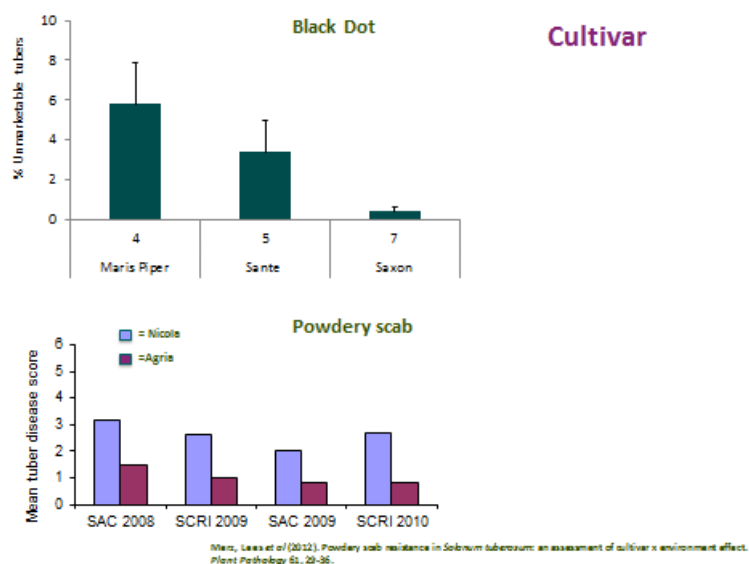


- Associated environmental variables known
- Can study relationship between environment and infection/disease



What do the results mean?

- Control options



Control – black dot



- Over all trials, azoxystrobin reduced the percentage of tubers unmarketable due to black dot, from 26.7% to 14.6 % (Maris Piper), and from 12.9 % to 7.1 % (Sante).
- Longer crop duration (120-130 days from 50% emergence to harvest date) increases the risk of black dot developing, particularly where a high level of soil inoculum is present





Control - Powdery Scab

- Assess inoculum on seed and in soil
- Select fields and assign varieties accordingly
- Select a free-draining soil
- Avoid over-cultivation/compaction
- Avoid over-irrigation
- Consider chemical control
 - Fluazinam (Extension of Authorisation) in seed crops.



Disease risk assessment



- Soil test result → risk
- Variety grown → Reduce/increase risk
- Likely crop duration → Reduce/increase risk
- Irrigation → Increased risk
- Chemical control → Reduce risk (may depend on level of inoculum)
- Weather conditions ?



Managing the risk of black dot

HIGH RISK	FACTOR	ACTION	TECHNICAL
VARIETY	Check varietal resistance to black dot at www.dorfora.org.uk/ varieties, in the SLAB pocket guide, or contact your supplier	Although the majority of pea-pod varieties are susceptible to black dot, there are a number of moderately resistant varieties. Resistance ratings vary from 1 (very susceptible) to 5 (resistant). The most susceptible varieties should not be planted in fields known to be contaminated with black dot inoculum at high risk levels. Susceptible varieties include: Taurus, King Edward, Wawa, Piper and Portland (these have recently been replaced by Cabernet, Savoy, and Earth).	Varietal resistance can help reduce the incidence of black dot in Peas. Dorset Hutton have investigated control measures against black dot, comparing a range of susceptible varieties to a more resistant variety resulted in a significant reduction in black dot.
FIELD CONDITIONS	Perform a black dot soil test (this is particularly valuable on rented land with unknown field history). Otherwise, if field history is known, take this into account e.g. crops grown in short rotations with a history of black dot, presence of hard seed populations and high volunteer numbers may be high risk.	Soil borne contamination is the most important source of inoculum for disease. However, a soil test may not be a reliable indicator of the level of black dot inoculum in a field. A number of related factors are linked. Areas with the highest level of soil contamination can be avoided.	Figure 1: Impact of soil borne inoculum on black dot incidence.
SEED QUALITY	Avoid contaminated seed of more susceptible varieties and avoid planting contaminated seed in soils not contaminated with black dot	The relationship between the amount of black dot on seed and disease on daughter plants is complex. It is widely accepted that seed with high black dot levels negatively affect...	Using the soil test, the risk of disease can be split into three categories depending on level of soil inoculum. The risk of 1000 kg DMS/kg seed, reduced to 100-1000 kg DMS/kg seed, and high risk (>1000 kg DMS/kg seed). As part of a Peas Choice trial, 100 DMS were compared to soil contamination. 85% of seeds were found to be contaminated, with 30% having a moderate to high risk.
CROP DURATION	Harvest by early October or end duration in the ground according to susceptibility and maturity group	Harvesting early can reduce the amount of black dot on seed and disease on daughter plants to a certain extent. However, the relationship between the amount of black dot on seed and disease on daughter plants is complex. It is widely accepted that seed with high black dot levels negatively affect...	Figure 2: Effect of harvest on disease risk. Soil borne inoculum is high when a crop is harvested in 2015, 2016, and 2017. Soil borne inoculum is low when a crop is harvested in 2018.
IRRIGATION	Avoid over irrigation especially of susceptible varieties and long-duration crops	Black dot is a disease favoured by wet and warm soil conditions. Water and irrigation in growth, together at high levels of summer rainfall will increase black dot, particularly where a susceptible variety of a long duration crop is grown. Water should be taken into account when making decisions on irrigation. Irrigation should be avoided in early autumn, reducing initial inoculum and harvesting as early as possible.	Figure 3: Effect of irrigation on disease risk. Soil borne inoculum is high when a crop is irrigated in 2015, 2016, and 2017. Soil borne inoculum is low when a crop is irrigated in 2018.
TEMPERATURE	Rapid pull-down to holding temperature	Reduced disease from susceptibility, it is probable to reduce the long term risk of infection by cooling after harvest. For a 2.5°C and 10°C per day in cooling, temperature, the number of susceptible plants is reduced by 10% in 10 days. However, the number of plants is reduced by 10% in 10 days. However, the number of plants is reduced by 10% in 10 days.	Figure 4: Effect of cooling on disease risk. Soil borne inoculum is high when a crop is cooled in 2015, 2016, and 2017. Soil borne inoculum is low when a crop is cooled in 2018.

Bringing together research findings, commercial tests and disease control advice

Late Blight?

Populations – implications for:
Fungicidal control
Host resistance
Prediction/risk
Precision approaches

Current and Future



- Field/site based diagnostics. Vegetables and sugar beet?
- PCN model – temperature v infestation
- Late blight, populations, field-based risk, precision approaches
- FLN as indicators of soil health
- FLN mapping & targeting of nematicide
- Tramline erosion/run-off (spread of pathogens)
- Optical imaging to detect disease risk/map fields



Stuart Wale
Alex Hilton
Daan Kiezebrink



Jeff Peters
James Woodhall



Peter Gladders
Faye Ritchie

Tonya Wiechel, Dept. Primary Industries, Aus.
Leigh Sparrow, Tasmania
Andrew Pitman, Plant & Food Research, NZ
Jacquie van der Waals, U of Pretoria, SA



Crop Duration



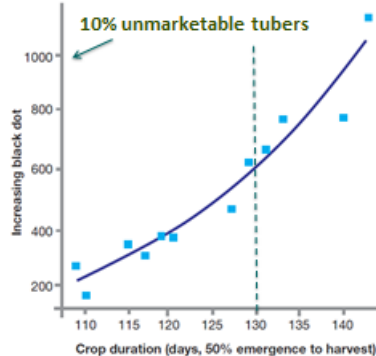
High Risk: Harvest date after early October or where crops are grown for a long duration (120-130 days from 50% emergence to harvest date)

Technical: Absolute date of harvest is less important than crop duration.

There is a good relationship between black dot development and crop duration (from 50% emergence to harvest)

The relationship is affected by maturity group – for susceptible maincrop varieties risk of economic loss is higher after 130 days

Crop Duration



Action: Harvest by early October or limit duration in ground according to susceptibility and maturity group

Dr Krishna Subbarao's presentation at Industry Workshop

**Broccoli-Mediated Pathogen and Disease
 Suppression in Vegetable Cropping Systems**

K. V. Subbarao

Department of Plant Pathology
 University of California, Davis



GROSS PRODUCTION VALUE

CATEGORIES	2013 TOTAL VALUE	2012 TOTAL VALUE*
Vegetable Crops	\$2,833,775,000	\$2,557,772,000
Fruit & Nuts	\$1,159,589,000	\$1,057,684,000
Nursery Crops	\$312,346,000	\$307,543,000
Livestock & Poultry	\$45,024,000	\$53,126,000
Field Crops	\$19,990,000	\$19,338,000
Seed Crops	\$8,803,000	\$8,550,000
Apiary	\$195,000	\$204,000
TOTALS	\$4,379,722,000	\$4,004,217,000

		1993	2003	2013
TOTAL OF MAJOR CROPS ABOVE	Acre	242,482	276,648	282,681
	Value	\$1,430,953,000	\$2,446,538,000	\$3,668,394,000
	CPI Adjusted	\$2,307,987,000	\$3,096,883,000	\$3,668,394,000

Monterey County's Ten Million Dollar Crops

CROPS	2013 CROP VALUE	2013 CROP RANKING	2012 CROP RANKING
Strawberry	\$869,488,000	1	1
Leaf Lettuce	\$659,646,000	2	2*
Head Lettuce	\$550,628,000	3	3
Broccoli	\$426,933,000	4	4
Celery	\$217,452,000	7	7
Misc. Vegetables	\$173,602,000	8	8
Cauliflower	\$163,319,000	9	10
Spinach	\$122,676,000	10	9
Mushrooms	\$71,534,000	11	12
Salad Products	\$70,577,000	12	13
Spring Mix	\$70,140,000	13	11
Artichokes	\$47,390,000	14	14
Cabbage	\$45,127,000	15	16
Raspberries	\$43,791,000	16	15





Background

- Extensive surveys revealed the high degree of susceptibility of cauliflower and near-immunity of broccoli to *Verticillium* wilt.
- *Verticillium dahliae* isolates from crucifer crops were weakly pathogenic on broccoli in greenhouse tests, and others were non-pathogenic.
- Dry crucifer residues are better than fresh residue.

Reaction of broccoli to *Verticillium dahliae* from various hosts

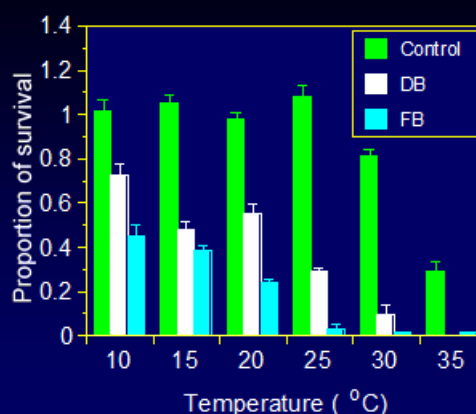
Isolate	Disease severity	Dry weight (g)	
		Root	Shoot
Cauliflower	1	2.31	4.07
Chili pepper	0	2.57	3.90
Artichoke	1	2.28	3.58
Cabbage	1	2.18	4.34
Lettuce	0	2.52	4.02
Strawberry	0	2.41	3.60
Bell pepper	0	2.50	4.23
Eggplant	0	2.57	3.61
Tomato	1	2.14	4.06
Watermelon	0	1.90	3.75
Mint	1	2.29	4.22
Potato	1	2.79	3.62
Cotton	0	3.03	4.17
Alfalfa (<i>albo-atrum</i>)	0	2.61	3.97
Control	0	2.71	4.42
LSD ($P \leq 0.05$)		0.55	0.58
Cauliflower	5	1.80	2.25

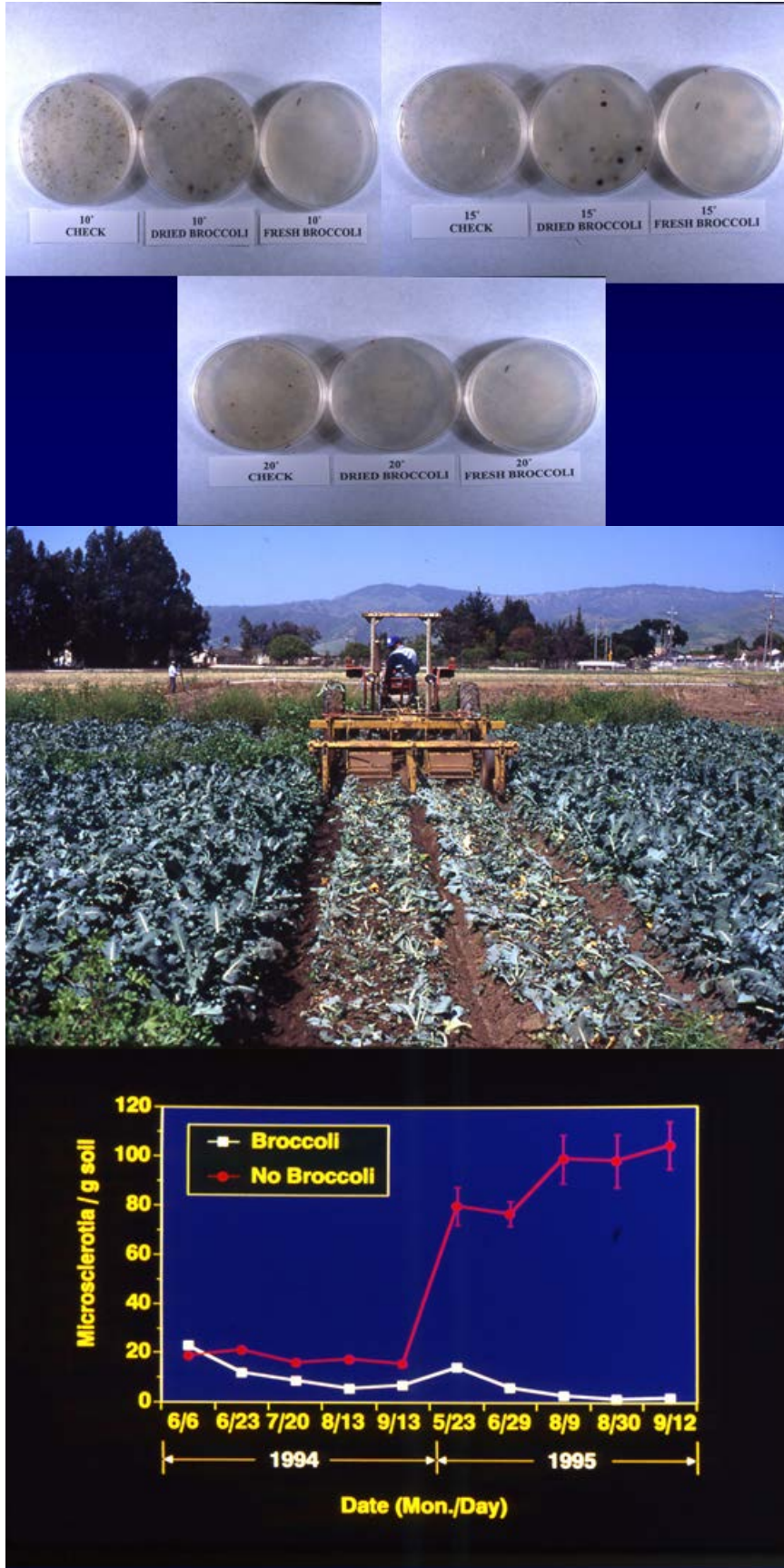
Questions

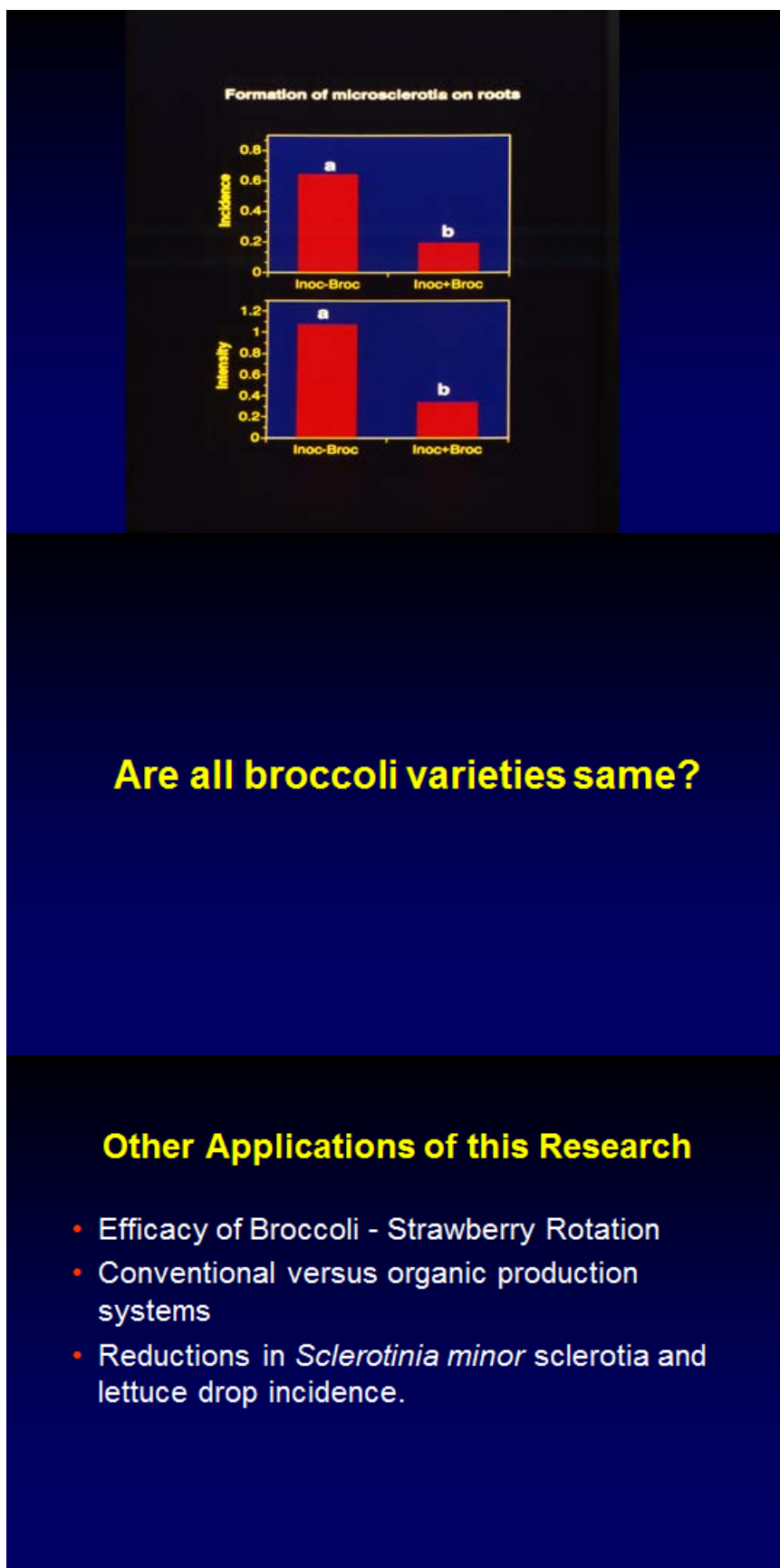
- When should the broccoli crop be planted and residue incorporated?
- Is there a specific temperature at which propagule reduction by broccoli is maximized?
- Is dry broccoli better than fresh broccoli?

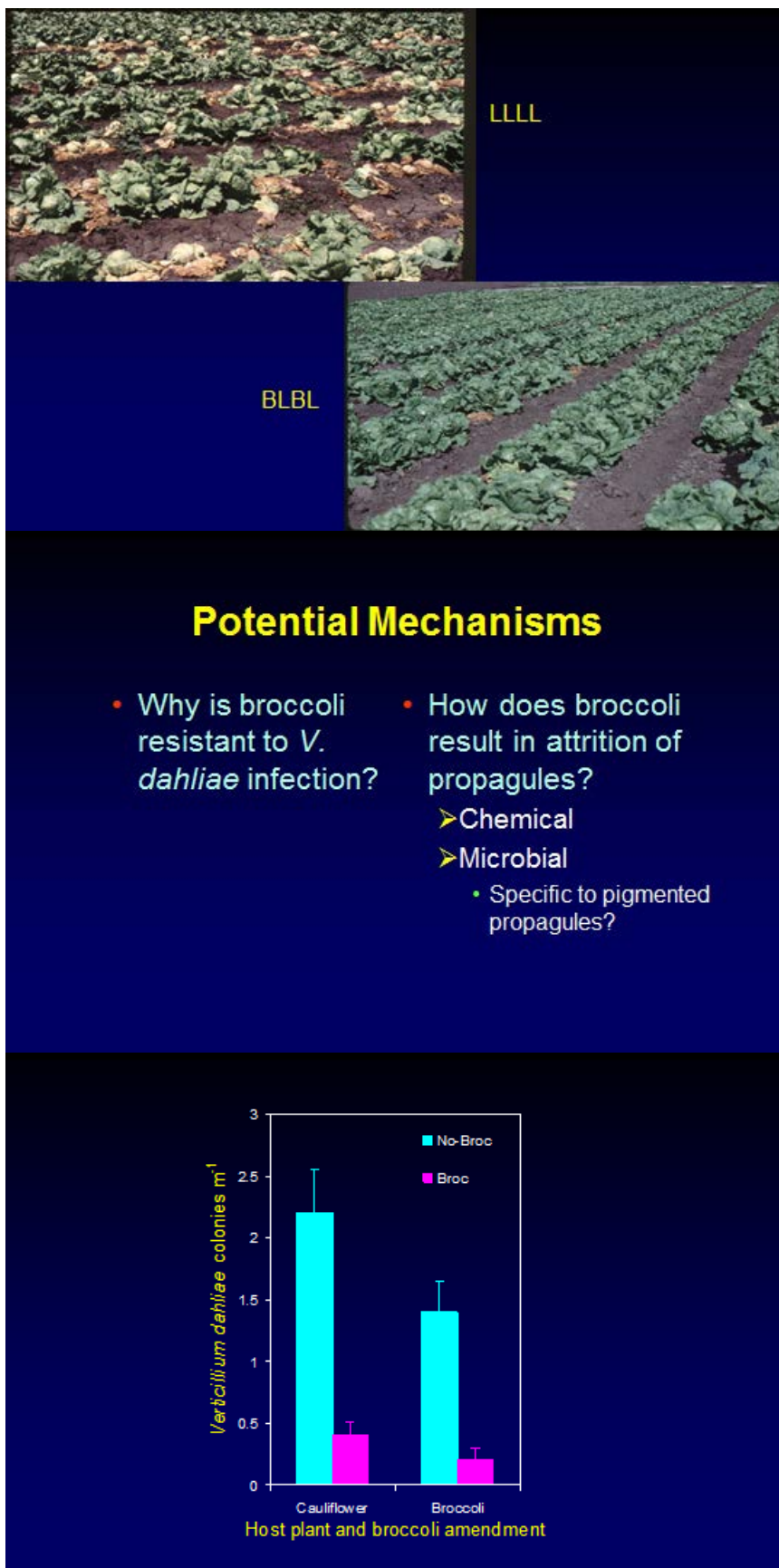
Methods

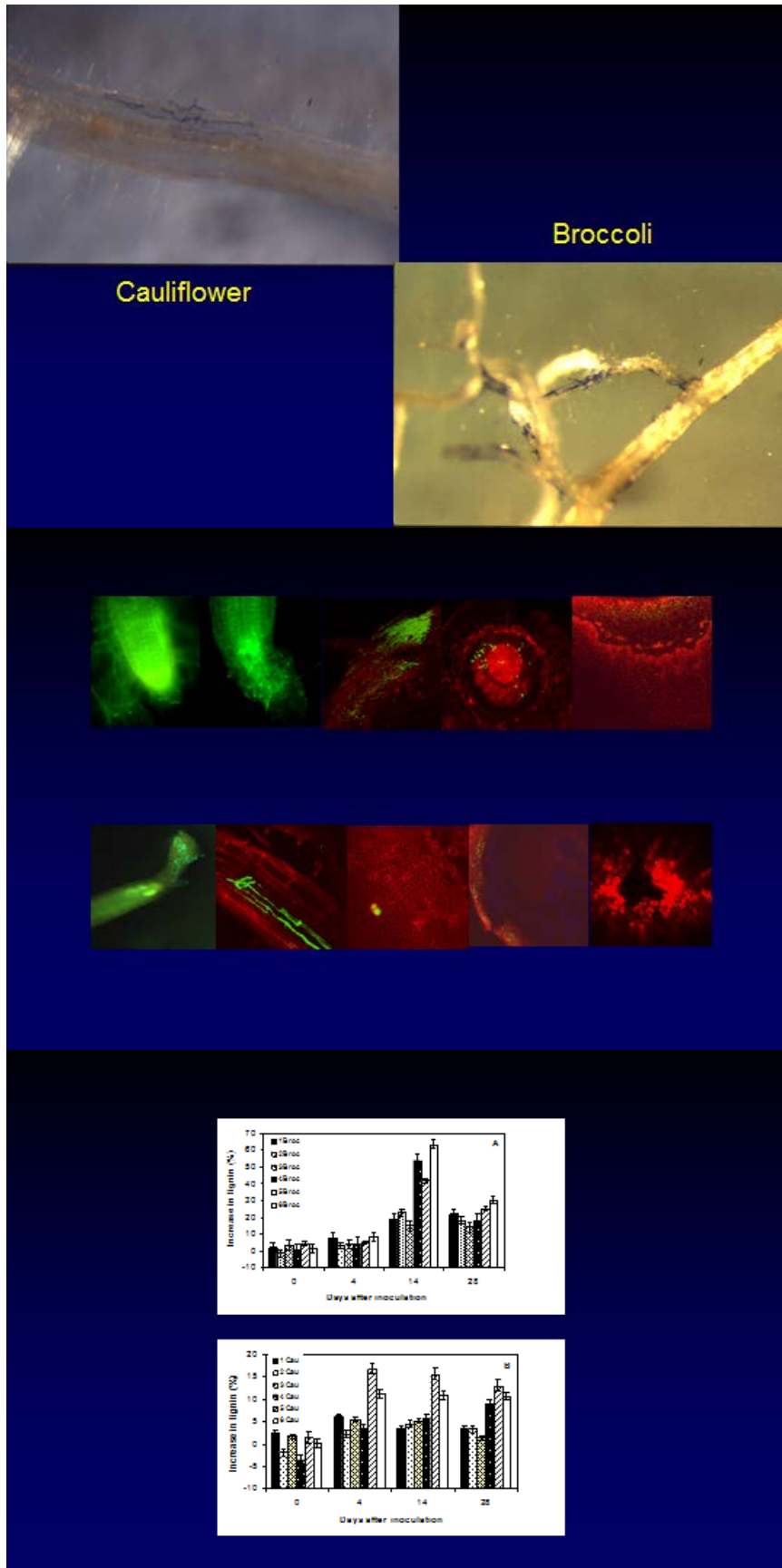
- **Soil:** Fields 1 & 2
- **Treatments:** Fresh (8%), dry, and no broccoli in 25 g dry soil.
- **Incubation:** 10, 15, 20, 25, 30, & 35 C
- **Assay:** After 45 days incubation using the Anderson sampler technique.
- In a parallel set of experiments, soil sampled after 15, 30, and 45 days incubation and assayed.











Summary of cover crop biomass and equivalent metam sodium content

Treatment	Biomass (T/A)		N (lb/A)		Metam sodium Eq (gal/A) – 2005
	2004	2005	2004	2005	
Broccoli	2.08	1.84	132.8	71.0	0.11
White Mustard	2.51	1.78	194.8	130.8	2.28
Indian Mustard	2.23	1.33	199.7	120.5	1.69

Labelled rates of metam sodium 30 -75 gal/A

Soil Microbiological Changes

Significant differences between broccoli and cauliflower.

Broccoli residue incorporation results in 100-fold increases in bacteria and 1000-fold increase in actinomycetes.

Not only these changes are quantitative but also qualitative. The diversity of these groups is highest in broccoli-amended soils compared with cauliflower-residue amended soils.

Bacterial diversity - Broccoli

- *Arthrobacter saperdae*
- *A. barkeri*
- *A. oxydans*
- *A. ramosus*
- *A. viscosus*
- *Aureobacterium esteraromaticum*
- *Bacillus circulans*
- *B. atrocyaneus*
- *B. brevis*
- *B. lentus*
- *B. pantothenicus*
- *B. psychrophilus*
- *B. pumilus*
- *Micrococcus lylae*
- *M. kristinae*
- *M. roseus*
- *Pseudomonas florescens*
- *Stephylococcus epidermis*
- *S. cohnii*
- *Stenotrophomonas maltophilia*

Bacterial diversity – non-broccoli

- *Bacillus chlororaphis*
- *B. laterosprus*
- *B. linens*
- *B. psychrophilus*
- *Curtobacterium inologenes*
- *C. flaccumfaciens*
- *Micrococcus halobius*
- *Pseudomonas putida*

Myxobacteria

- Gram negative, gliding bacteria
- Produce swarms on nutrient poor media
- Produce characteristic fruiting bodies with dormant myxospores
- Lytic activity on microorganisms and nematodes

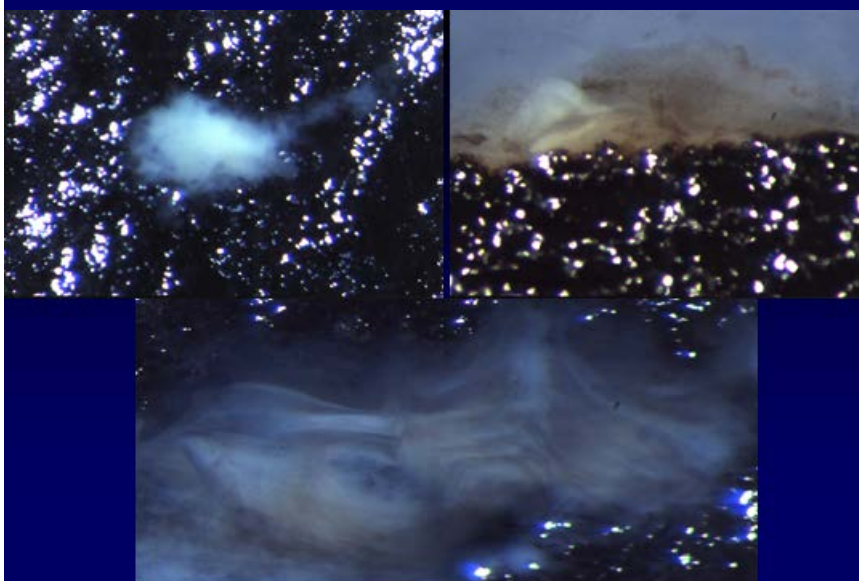


Myxobacteria

- *Myxococcus coralloides*
- *M. fulvus*
- *M. virescens*
- *M. xanthus* (2)
- *M. stipitatus*
- *M. flavascense*

Benefits of Melanin

- Abiotic Stress
UV radiation, temperature extremes, desiccation, free radicals, metal toxicity
- Biotic Stress
Soil microbial/host plant origin hydrolytic enzymes and free radicals
- Pathogenicity
Appressorial penetration, virulence factor?





CONCLUSION

The ASDS8 conference was a success. Informal feedback from participants and invited speakers praised the quality of the meeting and the opportunities it raised.

The invited speakers were well received by both the scientific community and industry at the workshop. In particular, Prof Subbarao's presentation on the limitations of biofumigants, but success with changes in farming systems was illuminating given current interest in biofumigants.

TECHNOLOGY TRANSFER

SCIENTIFIC CONFERENCE PAPERS

The ASDS8 conference featured 50 oral and 24 poster presentations relating to soil-borne disease (see attached program).

INDUSTRY & MEDIA PUBLICATIONS

Two articles about the keynote speaker featured within Vegetables Australia and Potatoes Australia.

One article interviewed Prof. Krishna Subbarao (UC Davis, USA) and focussed on his work with *Verticillium dahliae* and *Sclerotinia* spp. in Californian vegetable production systems. These diseases are very topical for Australian producers. His outcomes highlighted how changes in farming systems had dramatic effects on disease incidence.

The other article interviewed Dr Alison Lees, and focussed on her insights into potato diseases, and collaborations with Australian researchers within the APRP#2 program.

- “International research on show in Tasmania”. Vegetables Australia Jan/Feb 2015: 38-39.
- “International researcher shares knowledge with Tassie growers”. Potatoes Australia Dec/Jan 2015: 18-19

Interviews during the conference by Dr Calum Wilson and Prof Subbarao were featured on radio during the ABC country hour.

An article on the ASDS conference appeared in the Tasmanian Country newspaper featuring interviews from Dr Jason Scott, Dr Robert Tegg and prof Krishna Subbarao.

INDUSTRY & PEER PRESENTATIONS

The industry workshop held at the Forthside Research Farm provided an opportunity for the Tasmanian vegetable sector to engage with the key note speakers.

The workshop featured presentations from both speakers followed by a formal question time and lunch with informal discussions.

PROJECT RECOMENDATIONS

The ASDS meetings are of critical importance to Australian horticulture. they provides one of the few opportunities for Australasian specialists in soil-borne disease to meet and discuss the latest developments in this field. There are many unanswered questions relating to control of soil-borne disease. This is reflected in the proportion of R, D&E projects in this area. With diminishing resources invested in soil-borne disease research it is imperative that those active in the area share information and form collaborations to use these resources in the most efficient manner. ASDS provide a perfect venue for such developments.

We recommend HIA consider sponsorship opportunities for future iterations of this important meeting (the next to be held in Hamner Springs, New Zealand)